# Synthesis of 3- and 5'-substituted flavone-8-carboxylic acids as 'three-armed' leukotriene $CysLT_I$ receptor antagonists

Mariël E. Zwaagstra, Ronald E.M. Korthouwer, Henk Timmerman, Ming-Qiang Zhang\*

Division of Medicinal Chemistry, Leiden-Amsterdam Center for Drug Research, Vrije Universiteit, De Boelelaan 1083, 1081 HV Amsterdam, The Netherlands

(Received 12 June 1997; accepted 10 September 1997)

**Abstract** – Molecular modelling of leukotriene  $CysLT_I$  receptor antagonists have suggested that in addition to the two binding sites for a lipophilic and an acidic group, the receptor has a 'third pocket' to accommodate 'three-armed' ligands such as montelukast 1. Based on the most rigid  $CysLT_I$  receptor antagonist 3'-[2-(2-quinolinyl)ethenyl]flavone-8-carboxylic acid 2, we have synthesised 3- and 5'-substituted flavone derivatives to probe this additional binding pocket. Introduction of large substituents, e.g. 2-quinolinyl-methoxy, to the C5' position of the flavone skeleton abolished the  $CysLT_I$  receptor affinity whereas the same modification at the C3 position yielded a potent  $CysLT_I$  antagonist. This observation implies that the third binding pocket of the receptor has considerable steric tolerance, probably corresponding to the substituents at C3 of the flavone skeleton. Further modification by introducing a C3 substituent containing a basic nitrogen resulted in compound **6g** with potent  $H_I$  antihistaminic activity although the  $CysLT_I$  antagonistic activity was much reduced. Further study on the  $CysLT_I$  receptor recognition of three-armed antagonists may facilitate the design of more effective antiasthmatic agents, e.g. dual antagonists of histamine  $H_I$  and leukotriene  $CysLT_I$  receptors. © Elsevier, Paris

antiasthmatic agents / histamine H<sub>1</sub> antagonists / leukotriene D<sub>4</sub> antagonists / molecular modelling

### 1. Introduction

Over the past decade evidence has accumulated showing that leukotrienes play an important role in the pathogenesis of asthma [1–6]. Structurally leukotrienes are divided into two groups: the hydroxyleukotriene LTB<sub>4</sub> and the cysteinyl leukotrienes (CysLTs) LTC<sub>4</sub>, LTD<sub>4</sub> and LTE<sub>4</sub>. CysLTs appear to be more important mediators than LTB<sub>4</sub> in asthma, especially as bronchoconstrictors. CysLTs also cause mucus secretion, increase vascular permeability, and are involved in the inflammation and hyperresponsiveness of human airways. Asthmatic subjects are more sensitive to the actions of CysLTs [7] and elevated levels of these endogenous substances have been detected during asthmatic attacks [8, 9]. Recent clinical success with CysLT antagonists, e.g. zafirlukast, pranlukast, montelukast [3, 10-12], represents the most significant advance in the acquisition of antiasthmatic agents for the last 20 years.

CysLTs produce their effects by stimulation of specific membrane receptors belonging to the G-protein-coupled superfamily [13, 14]. Two subtypes have been identified to bind specifically to CysLTs:  $CysLT_1$  for LTD<sub>4</sub> and LTE<sub>4</sub>, and  $CysLT_2$  for LTC<sub>4</sub>. Stimulation of the  $CysLT_1$  receptor leads to the characteristic symptoms of asthma whereas the consequences of selective activation of the  $CysLT_2$  receptor remain to be clarified.

In our search for effective antiasthmatic agents we became interested in  $CysLT_1$  receptor antagonists [15, 16]. Our attempts to define antagonist recognition of CysLT<sub>1</sub> receptors started with the synthesis of a series of carboxylated flavonoids, which resulted in the discovery of two new leads 4-hydroxy-3-[1-oxo-2propenyl-3-(2-quinolinyl)]benzoic acid and 6-bromo-8-carboxy-4-oxo-2-(2-quinolinyl)-1-4H-benzopyran with weak CysLT<sub>1</sub> receptor affinity [16]. Further modification afforded a series of carboxylated chalcones [17] and flavones [18] with potent  $CysLT_1$  antagonistic activity. The flavone series contains the most rigid CvsLT<sub>1</sub> receptor antagonists known to date [18], for example 3'-(2-(2-quinolinyl)ethenyl)flavone-8carboxylic acid (2, VUF 5017). Consequently we have developed a 3D model of CysLT<sub>1</sub> antagonists using 2 as templates for the superimposition of various known  $CysLT_1$  antagonists [18].

<sup>\*</sup>Correspondence and reprints Current address: Scientific Development Group, Department of Medicinal Chemistry, Organon Laboratories Ltd., Newhouse, ML1 5SH, Scotland, U.K.

Based on this model we have shown that in addition to the two characteristic binding sites for a lipophilic group and an acidic function, respectively, there is a third pocket in the receptor cavity to accommodate the 'third-arm' of antagonists such as montelukast 1 and sulukast [10] (figure 1). Computer-aided superimposition (figure 1) of 1 and 2 revealed that this additional binding pocket might correspond to a substituent at either the C3 or the C5' position of the flavone skeleton. We therefore prepared derivatives of 2 by introducing various substituents at the C3 and C5' positions in order to probe this third binding pocket of the  $CysLT_1$  receptor. In this paper we wish to report the synthesis and in vitro activity of these new flavone derivatives containing 3- or 5'-substituents. Preliminary attempts to design dual antagonists of histamine H<sub>1</sub> and leukotriene CysLT<sub>1</sub> receptors based on the above modification is also included. It has been proposed that such dual antagonists may be more effective antiasthmatic agents than selective  $CysLT_i$  antagonists [15, 19].

# 2. Chemistry

For the synthesis of 'three-armed' flavones it would be most convenient to introduce a wide range of substituents at a late stage. Since alkylation of 3- or 5'-hydroxyflavones provides a facile route to various substituents having different physicochemical properties, we chose 3- and 5'-alkyloxy derivatives as our targets. The 3-hydroxyflavones 4, the key intermediates for a wide range of 3-substituted flavone derivatives, were obtained from the corresponding chalcones 3 [17] in a one-step procedure (Algar–Flynn–Oyamada oxidation [21, 22]) (figure 2) in about 50% yield. Alkylation of 4 with an appropriate alkyl halide and the subsequent hydrolysis of the resulting esters 5 afforded the 3-substituted flavones 6.

Although it seemed straightforward, alkylation of 4 in some cases may be troublesome. For instance, alkylation with 1,3-dibromopropane, 1-bromo-3-chloropropane or 3-bromo ethyl propionate gave unidentified mixtures of products. Hydrolysis of the esters 5

Figure 1. Correspondence of the 'third arms' of  $CysLT_i$  receptor antagonists.

Figure 2. Synthesis of 3-substituted flavones 6.

generally proceeded smoothly under the influence of sodium- or potassium hydroxide in a mixture of water and ethanol. In some cases THF was added to improve the solubility of the reactants. Target compounds 6 were obtained in 10–15% overall yield.

Compound **6g** was synthesised by reductive alkylation [23] of the primary amine **6f** which was obtained from the phthalimide **6e** with hydrazine (*figure 3*).

For the synthesis of 5'-substituted flavones, 5'-hydroxy-3'-(2-quinolinylmethoxy)flavone was considered as a useful precursor. The synthetic approach depicted in *figure 4* was used for obtaining this compound [24–26]. This route involved the coupling of 7 and acetophenone 8 to form the 1,3-diketone 9. This intermediate was cyclodehydrated to flavone 10 which upon demethylation with boron tribromide in dichloromethane afforded 3',5'-dihydroxyflavone 11 in good yield. Aluminium chloride or bromide were found not suitable for the demethylation of 10. Esterification of 11 provided compound 12.

Alkylation of 12 in the presence of one equivalent 2-chloromethylquinoline resulted in the isolation of

the dialkylated product 13 only, instead of the desired mono-substituted flavone. Apparently, the increased electron-donating effect of the 2-quinolinylmethoxy group in the monoalkylated flavone resulted in an increased reactivity of the second hydroxyl group to such an extent that the monoalkylated flavone is much more reactive towards the alkylating agent than the dihydroxyflavone 12. Hydrolysis of 13 in a mixture of DMF-water in the presence of potassium hydroxide yielded 3',5'-bis(2-quinolinemethoxy) 14.

#### 3. Results and discussion

The CysLT<sub>1</sub> receptor affinities of 3- and 5'-substituted flavones are shown in table I, together with the receptor affinities of the reference compounds VUF 4918, VUF 4871 and VUF 5165 [18]. Compared with VUF 4918, the 3-hydroxyl analogue 4a showed a 25-fold decreased affinity and the 3-carboxymethoxy analogue 6a had even lower affinity. This seems to suggest that small and hydrophilic substituents are not

Figure 3. Synthesis of 6g.

Figure 4. Synthesis of 5'-substituted flavone 14.

**Table I.** CysLT<sub>1</sub> receptor affinity of 3- and 5'-substituted flavones.

Compound	Sub. pos.	$R_1$	$R_2$	$R_3$	$K_{\rm D}$ (nM) <sup>a</sup> , or (% inhib. at 10 <sup>-5</sup> M
VUF 4918	3'	Н	Н	Н	17 ± 3
VUF 4871	4'	Н	Н	Н	$526 \pm 88$
VUF 5165	3'	Br	Н	H'	$31 \pm 9$
4a	3'	Н	ОН	Н	$452 \pm 97$
4b	3'	Br	ОН	Н	$38 \pm 9$
4c	4'	Н	ОН	Н	$805 \pm 130$
6a	3'	Н	OCH <sub>2</sub> CO <sub>2</sub> H	Н	$639 \pm 78$
6b	3'	Br	OCH <sub>2</sub> CO <sub>2</sub> H	Н	$40 \pm 7$
6c	3'	Br	OCH <sub>2</sub> CH <sub>2</sub> CH <sub>3</sub>	Н	$44 \pm 2$
6d	3'	Br	2-quinolinylmethoxy	Н	$63 \pm 3$
6e	4'	Н	O(CH <sub>2</sub> ) <sub>2</sub> phthalimide	Н	$916 \pm 166$
6g	4'	Н	$O(CH_2)_2NMe_2$	Н	$4020 \pm 430$
14	3'	Br	H	2-quinolinylmethoxy	(5%)

<sup>&</sup>lt;sup>a</sup>The affinity data were obtained from binding assays in guinea pig lung membranes versus [ ${}^{3}H$ ]LTD<sub>4</sub>. The data are means  $\pm$  S.E.M. of three determinations.

favourable at this position. However, when compared with VUF 4871, the corresponding 3-hydroxyl analogue **4c** was similarly active, maybe due to the low sensitivity of less active compounds towards modification (VUF 4817 and **4c** are much less active than their 3'-(2-quinolinemethoxy) counterparts VUF 4918 and **4a**).

Among the 6-bromo derivatives, the sensitivity towards 3-hydroxylation was virtually invisible and the 3-hydroxyflavone **4b** was as active as the non-hydroxy VUF 5165. This insensitivity towards substitution at C3 continued with carboxymethoxy **6b**, *n*-propoxy **6c** and 2-quinolinemethoxy **6d**. When compared with the non-brominated analogues VUF 4918, **4a** and **6a**, this insensitivity towards substitution at C3 seems to result from the bromine atom at position 6. Whether this is an electronic or lipophilic factor remains to be further investigated.

The most interesting structure–activity relationship within this series are the different effects of the introduction of a 2-quinolinemethoxy group to the C3 and C5' positions. The 3-substituted compound **6d** showed an affinity value ( $K_D$ ) of 63 nM but the 5'-substituted compound **14** was virtually inactive. This observation implies that the third binding pocket of the  $CysLT_I$  receptor corresponds to the substituent at position 3 of the flavone skeleton and has considerable steric tolerance.

In addition to the CysLT<sub>1</sub> receptor affinity discussed above, we have noticed that 6g possessed some structural characteristics of histamine H<sub>1</sub> antagonists. Indeed the compound exhibited high affinity to histamine  $H_1$  receptor with a  $K_D$  value of 10 nM versus [3H]mepyramine in guinea pig lung membrane. Histamine H<sub>1</sub>-receptor antagonists are very useful therapeutic agents for many allergic disorders [27]. Due to the complementary actions of CysLTs and histamine during inflammatory and allergic responses [28–30], compounds with dual antagonism of  $CysLT_1$ and H<sub>1</sub> receptors may be more effective than any single mediator antagonists in the treatment of asthma [15, 19]. Further modification of 6g towards a better balanced dual antagonism of H<sub>1</sub> and CysLT<sub>1</sub> receptors may provide eventually more effective antiasthmatic agents.

#### 4. Conclusions

Based on the superimposition of 1 (montelukast) and 2 as incorporated in our 3D  $CysLT_1$  antagonist model [18], we have suggested that a 'third arm' may be introduced into the flavone skeleton. Introduction of a 2-quinolinemethoxy substituent revealed that the antagonist binding site differentiated the substitutions at the 3- and 5'-positions, with the additional inter-

action point probably locating at a place corresponding to that of the C3 substituent of the flavone skeleton. Further modification on the C3 position, e.g. introduction of a nitrogen-containing group, may result in compounds with better antiasthmatic profiles, e.g. dual antagonism of histamine  $H_1$  and leukotriene  $CysLT_1$  receptors [15, 19].

# 5. Experimental protocols

# 5.1. Pharmacology

5.1.1. Radioligand binding assay with [3H]LTD<sub>4</sub>

The method is very similar to that described previously [15]. Briefly, a mixture of total volume of 0.3 mL containing 0.2 nM [3H]LTD<sub>4</sub>, guinea-pig lung membrane fractions (± 170 µg/mL) and the testing compound in a 10 mM piperazine-N,N'-bis(2ethanesulphonic) acid buffer (pH 7.5) was incubated at 22 °C for 30 min. The piperazine-N,N'-bis(2-ethanesulphonic) acid buffer contains 10 mM CaCl<sub>2</sub>, 10 mM MgCl<sub>2</sub>, 50 mM NaCl, 2 mM cysteine and 2 mM glycine. The reaction was terminated by the addition of 5 mL ice-cold Tris-HCl/NaCl buffer (10 mM/100 mM, pH 7.5). The mixture was immediately filtered under vacuum (Whatman GF/C filters) and the filters were washed once with 20 mL ice-cold buffer. The retained radioactivity was determined by a liquid scintillation counter. In the saturation experiment, 2 µM LTD<sub>4</sub> was used to define the non-specific binding. A single, saturable binding site with  $B_{\text{max}} = 988$  fmol/mg protein was found from saturation experiments. The  $K_D$  of [3H]LTD<sub>4</sub> was established to be 2.16 x 10-10 M and no cooperativity was detected when the data were analysed by Hill plots (slope = 0.99).

5.1.2. In vitro inhibition of [<sup>3</sup>H]mepyramine binding to guineapig lung membranes

The method is based on that described previously [31]. Briefly, a mixture of a total volume of 1.0 mL containing 0.5 nM [³H]mepyramine (specific activity 21 Ci/mmol), guinea pig lung membrane proteins (± 370 mg/mL) and the testing compound in 50 mM Na–K phosphate buffer (pH 7.5) was incubated at 37 °C for 30 min. The reaction was stopped by the addition of 5 mL ice-cold phosphate buffer and followed by immediate filtration through Whatman GF/C filters. The filters were washed twice with about 20 mL cold buffer. The retained radioactivity was determined by a liquid scintillation counter after addition of 5 mL scintillation liquid.

In the saturation experiment,  $10^{-4}$  M R-(-)-dimethindene was used to define the non-specific binding. A single, saturable binding site with  $B_{\text{max}} = 278 \pm 24$  fmol/mg protein was found from the saturation experiment. The  $K_{\rm D}$  of [3H]mepyramine was found to be  $3.30 \pm 0.26 \times 10^{-9}$  M and no cooperativity was detected when the data were analysed by Hill plots (slope = 1.005).

#### 5.2. Chemistry

<sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra were recorded on a Bruker AC 200 (<sup>1</sup>H-NMR: 200.1 MHz, <sup>13</sup>C-NMR: 50.29 MHz) or a Bruker 400 MSL spectrometer (<sup>1</sup>H-NMR: 400.1 MHz, <sup>13</sup>C-NMR: 100.63 MHz). <sup>1</sup>H-NMR chemical shifts (δ) are reported in ppm relative to CDCl<sub>3</sub> ( $\delta$  = 7.25 ppm) or DMSO- $d_5$  ( $\delta$  = 2.5 ppm). <sup>13</sup>C-NMR chemical shifts (δ) are reported in ppm relative to CDCl<sub>3</sub> ( $\delta$  = 77.0 ppm) or DMSO- $d_6$  ( $\delta$  =

39.5 ppm). Coupling constants are given in Hz. 2D-NMR (H–H and C–H-COSY) techniques were frequently used to support the interpretation of the 1D spectra. NOE-difference spectra were recorded on a Bruker 400 MSL spectrometer. The multiplicity of the carbon signals was determined by DEPT- or APT-spectra, or by a combination of a normal decoupled carbon spectrum in combination with a CH-correlation. The symbols used are (p) for primary, (s) for secondary, (t) for tertiary and (q) for quaternary carbon signals. The spectra were collected at room temperature at 200 MHz, unless stated otherwise.

FAB (HRMS) measurements were performed on a Finnigan MAT 90 spectrometer equipped with a WATV Cs ion gun, operated at a beam current of approximately 2  $\mu$ A at 25 kV. High-resolution mass spectra were recorded on a Finnigan MAT-90. Melting points were measured on a Mettler FP-5 + FP-52 apparatus equipped with a microscope and are uncorrected.

Starting materials were commercially available. All end products had elemental analysis within 0.4% of theoretical values. However, compounds obtained in too small quantities to perform the elemental analysis or having an elemental analysis slightly outside of this range were found to be pure by both spectroscopic and chromatographic criteria.

5.2.1. General procedure for the synthesis of 3-hydroxyflavones
 The 2'-hydroxy-(2-quinolinylmethoxy)chalcones 3 [17]
 (1.2 mmol) were dissolved in 35 mL methanol and 2 mL 16%
 KOH in water was added. The solution was cooled on ice and
 2.0 mL 15% H<sub>2</sub>O<sub>2</sub> was added dropwise. The reaction mixture was stored at 4 °C for 24 h and acidified with 1 N HCl. The precipitate was filtered and recrystallised from DMF to yield the pure products.

5.2.1.1. 3-Hydroxy-3'-(2-quinolinylmethoxy)flavone-8-carboxylic acid 4a: Mp 272.2–274.2 °C, yield 13%. Recrystallisation from DMF/ethanol.  $^1\text{H}$ -NMR (400 MHz, DMSO): δ 5.45 (s, 2H, CH<sub>2</sub>O), 7.23 (dd, 1H,  $^3J$  = 8.1 Hz,  $^4J$  = 2.2 Hz, H4'), 7.50 (t, 1H,  $^3J$  = 8.1 Hz, H5'), 7.55 (t, 1H,  $^3J$  = 7.7 Hz, H6), 7.61–7.65 (m, 1H, H6-quinoline), 7.76 (d, 1H,  $^3J$  = 8.5 Hz, H3-quinoline), 7.78–7.82 (m, 1H, H7-quinoline), 8.02 (d, 1H,  $^3J$  = 8.1 Hz, H5-quinoline), 8.05 (d, 1H,  $^3J$  = 8.5 Hz, H8-quinoline), 8.10 (d, 1H,  $^3J$  = 7.9 Hz, H6'), 8.20 (s, 1H, H2'), 8.30 (d, 1H,  $^3J$  = 7.4 Hz, H5/7), 8.35 (d, 1H,  $^3J$  = 7.3 Hz, H5/7), 8.45 (d, 1H,  $^3J$  = 8.5 Hz, H4-quinoline), 9.97 (br s, 1H, 3-OH).  $^{13}\text{C-NMR}$  (50 MHz, DMSO): δ 70.72 (s), 113.63 (t), 116.20 (t), 119.72 (t), 120.76 (t), 122.04 (q), 123.79 (t), 126.48 (t), 127.02 (q), 127.79 (t), 128.32 (t), 129.33 (t), 129.57 (t), 129.74 (t), 132.58 (q), 135.95 (t), 136.96 (t), 139.32 (q), 144.44 (q), 144.57 (q), 146.69 (q), 152.42 (q), 157.01 (q), 158.03 (q), 165.15 (q), 172.50 (q). MH+ 440 (base peak) for C<sub>26</sub>H<sub>17</sub>NO<sub>6</sub>. Anal. C<sub>26</sub>H<sub>17</sub>NO<sub>6</sub>·NaCl: C, H, N.

5.2.1.2. 6-Bromo-3-hydroxy-3'-(2-quinolinylmethoxy)flavone-8-carboxylic acid 4b: Mp 293.2–294.3 °C, yield 45%. Recrystallisation from DMF/ethanol. <sup>1</sup>H-NMR (200 MHz, DMSO):  $\delta$  5.44 (s, 2H, CH<sub>2</sub>O), 7.19–7.22 (m, 1H, H4'), 7.49 (t, 1H,  $^3J$  = 8.1 Hz, H5'), 7.59–7.65 (m, 1H, H6-quinoline), 7.75 (d, 1H,  $^3J$  = 8.4 Hz, H3-quinoline), 7.77–7.83 (m, 1H, H7-quinoline), 7.99–8.09 (m, 3H, H5-quinoline, H8-quinoline, H6'), 8.16 (s, 1H, H2'), 8.32 (d, 1H,  $^4J$  = 2.5 Hz, H5/7), 8.38 (d, 1H,  $^4J$  = 2.5 Hz, H5/7), 8.44 (d, 1H,  $^3J$  = 8.5 Hz, H4-quinoline), 10.18 (br s, 1H, 3-OH). <sup>13</sup>C-NMR (50 MHz, DMSO):  $\delta$  70.71 (s), 113.64 (t), 115.65 (q), 116.36 (t), 119.67 (t), 120.77 (t), 123.67 (q), 123.77 (q), 126.45 (t), 127.00 (q), 127.76 (t), 128.34 (t), 129.60 (t), 129.70 (t), 130.86 (t), 132.31 (q), 136.89

(t), 137.66 (t), 139.49 (q), 144.92 (q), 146.71 (q), 151.28 (q), 156.97 (q), 158.00 (q), 163.87 (q), 171.38 (q). MH+ 519 (base peak for  $C_{26}H_{16}BrNO_6$ . Anal.  $C_{26}H_{16}BrNO_6$ -0.5HCl: C, H, N.

5.2.1.3. 3-Hydroxy-4'-[(2-quinolinyl)methoxy]flavone-8-carboxylic acid 4c: Mp 226.5–227.8 °C, yield 49%. ¹H-NMR (400 MHz, DMSO):  $\delta$  5.47 (s, 2H, CH<sub>2</sub>O), 7.24 (d, 2H, H3', H5'), 7.30 (t, 1H,  $^3J$  = 7.6 Hz, H6), 7.61–7.64 (m, 1H, H6-quinoline), 7.72 (d, 1H,  $^3J$  = 8.5 Hz, H3-quinoline), 7.78–7.82 (m, 2H, H7-quinoline, H5/7), 7.94 (dd, 1H,  $^3J$  = 7.9 Hz,  $^4J$  = 1.6 Hz, H5/7), 8.01 (d, 1H,  $^3J$  = 8.4 Hz, H5-quinoline), 8.05 (d, 1H,  $^3J$  = 8.5 Hz, H8-quinoline), 8.44 (d, 1H,  $^3J$  = 8.5 Hz, H4-quinoline), 8.44 (d, 2H,  $^3J$  = 9.0 Hz, H2', H6').  $^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  70.68 (s), 114.21 (t), 114.58 (t), 119.47 (t), 121.33 (q), 123.34 (t), 123.62 (t), 124.46 (q), 126.44 (t), 127.00 (q), 127.77 (t), 128.37 (t), 129.67 (q), 132.42 (q), 132.69 (t), 136.92 (t), 137.61 (q), 144.84 (q), 146.75 (q), 151.31 (q), 156.98 (q), 158.00 (q), 166.71 (q), 171.94 (q), 172.63 (q).

5.2.2. General procedure for the preparation of 3-alkyloxy-flavone-8-carboxylic acids **6** 

A solution of 1 equiv 3-hydroxyflavones 4, 2.2 equiv of an appropriate alkylhalide and 2.2 equiv of potassium carbonate in DMF was stirred at 50–90 °C until completion of the reaction. The DMF was removed in vacuo, and water was added. The product was extracted with ethyl acetate, the combined organic layers were washed with water, 1 N NaOH and brine, and dried over sodium sulphate. Removal of the solvent yielded the crude product 5 which were further purified by crystallisation from ethanol.

The esters 5 were dissolved in a mixture of THF, methanol and 5% aq. LiOH (1:1:1). The reaction mixture was stirred at room temperature until completion of the hydrolysis (TLC: ethyl acetate). The reaction mixture was then acidified, and the precipitate was collected by filtration. The crude acids 6 were further purified by recrystallization from DMF.

5.2.2.1. 3-Hydroxycarbonylmethoxy-3'-(2-quinolinylmethoxy)flavone-8-carboxylic acid6a: Mp 230.8-232.6 °C (DMF/EtOH), yield 58%. <sup>1</sup>H-NMR (200 MHz, DMSO): δ 4.89 (s, 2H, CH<sub>2</sub>CO<sub>2</sub>H), 5.48 (s, 2H, CH<sub>2</sub>O), 7.27–7.31 (m, 1H, H4'), 7.50 (t, 1H,  $^{3}J = 8.0$  Hz, H5'), 7.54 (t, 1H,  $^{3}J = 7.9$  Hz, H6), 7.62–7.66 (m, 1H, H6-quinoline), 7.75 (d, 1H,  $^3J$  = 8.4 Hz, H3-quinoline), 7.78-7.82 (m, 1H, H7-quinoline), 8.03-8.06 (m, 3H, H5-quinoline, H8-quinoline, H6'), 8.20 (br s, 1H, H2'), 8.30 (d, 2H,  ${}^{3}J = 7.7$  Hz, H5, H7), 8.45 (d, 1H,  ${}^{3}J = 8.3$ , H4quinoline), 13.20 (br s, 1H, ArCO<sub>2</sub>H). 13C-NMR (50 MHz, DMSO): δ 67.32 (s), 70.76 (s), 114.50 (t), 117.76 (t), 119.66 (t), 121.37 (q), 121.60 (t), 124.00 (q), 124.41 (t), 126.46 (t), 127.01 (q), 127.78 (t), 128.34 (t), 129.27 (t), 129.55 (t), 129.73 (t), 131.57 (q), 136.11 (t), 136.90 (t), 139.21 (q), 146.74 (q), 152.42 (q), 153.00 (q), 157.04 (q), 157.93 (q), 165.02 (q), 169.81 (q), 173.24 (q). MS (m/z): [M + H]+ 498 (base peak) for C<sub>28</sub>H<sub>19</sub>NO<sub>8</sub>. Anal. C<sub>28</sub>H<sub>19</sub>NO<sub>8</sub>•0.25,HCl: C, H, N.

5.2.2.2. 6-Bromo-3-hydroxycarbonylmethoxy-3'-(2-quinolinylmethoxy)flavone-8-carboxylic acid **6b**: Mp 252.5–252.6 °C (DMF), yield 50%.  $^{1}$ H-NMR (200 MHz, DMSO):  $\delta$  4.88 (s, 2H, CH<sub>2</sub>CO<sub>2</sub>H), 5.49 (s, 2H, CH<sub>2</sub>O), 7.30 (d, 1H,  $^{3}$ *J* = 8.1 Hz, H4'), 7.51 (t, 1H,  $^{3}$ *J* = 8.0 Hz, H5'), 7.62–7.68 (m, 1H, H6-quinoline), 7.76–7.87 (m, 2H, H3-quinoline, H7-quinoline), 8.02–8.09 (m, 3H, H6', H5-quinoline, H8-quinoline), 8.19 (s, 1H, H2'), 8.35 (s, 2H, H5, H7), 8.49 (d, 1H,  $^{3}$ *J* = 8.6 Hz, H4-quinoline).  $^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  67.32 (s), 70.56 (s),

114.52 (t), 116.38 (q), 117.94 (t), 119.67 (t), 121.65 (t), 123.71 (q), 125.69 (q), 126.58 (t), 127.03 (q), 127.83 (t), 128.03 (t), 129.59 (t), 129.92 (t), 130.95 (t), 131.31 (q), 137.28 (t), 137.94 (t), 139.31 (q), 146.37 (q), 151.35 (q), 153.26 (q), 156.92 (q), 157.88 (q), 163.70 (q), 169.72 (q), 172.07 (q). Anal.  $C_{28}H_{18}BrNO_8 \cdot 0.25HCl:$  C, H N, Br.

5.2.2.3. 6-Bromo-3-propyloxy-3'-(2-quinolinylmethoxy)flavone-8-carboxylic acid **6c**: Mp 237.1–239.3 °C, yield 42%. ¹H-NMR (200 MHz, DMSO):  $\delta$  0.87 (t, 3H,  ${}^3J$  = 7.4 Hz, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.56–1.67 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 3.98 (t, 2H,  ${}^3J$  = 6.7 Hz, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 5.46 (s, 2H, CH<sub>2</sub>O), 7.29 (dd, 1H,  ${}^3J$  = 8.3 Hz,  ${}^4J$  = 2.6 Hz, H4'), 7.51 (t, 1H,  ${}^3J$  = 8.1 Hz, H5', 7.59–7.63 (m, 1H, H6-quinoline), 7.72 (d, 1H,  ${}^3J$  = 8.5 Hz, H3-quinoline), 7.67–7.70 (m, 1H, H7-quinoline), 792 (d, 1H,  ${}^3J$  = 8.0 Hz, H5-quinoline), 7.99–8.05 (m, 3H, H8-quinoline) H6', H2'), 8.32 (s, 2H, H5, H7), 8.44 (d, 1H,  ${}^3J$  = 8.6 Hz, H4-quinoline).  ${}^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  10.05 (p), 22.53 (s), 70.70 (s), 73.26 (s), 114.30 (t), 116.25 (q), 117.73 (t), 119.43 (t), 121.31 (t), 123.94 (q), 125.88 (q), 126.44 (t), 127.00 (q), 127.76 (t), 128.32 (t), 129.67 (t), 129.72 (t), 130.89 (t), 131.43 (q), 136.89 (t), 137.73 (q), 140.07 (q), 146.73 (q), 151.47 (q), 154.12 (q), 156.98 (q), 157.93 (q), 163.77 (q), 172.30 (q). MH+561 (base peak) for C<sub>29</sub>H<sub>23</sub>BrNO<sub>6</sub>. Anal. C<sub>29</sub>H<sub>23</sub>BrNO<sub>6</sub>: C, H, N, Br.

5.2.2.4. 6-Bromo-3,3'-di-(2-quinolinylmethoxy)flavone-8-carboxylic acid **6d**: Mp 229.6–229.7 °C, yield 51%. ¹H-NMR (200 MHz, DMSO):  $\delta$  5.34 (s, 2H, CH<sub>2</sub>O), 5.40 (s, 2H, CH<sub>2</sub>O), 7.26 (m, 1H, H4'), 7.44 (t, 1H,  $^3J=8.0$  Hz, H5'), 7.56–7.79 (m, 6H, 2 H6-quinoline, 2 H7-quinoline, 2 H3-quinoline), 7.91–8.03 (m, 6H, H6', H2', 2 H5-quinoline, 2 H8-quinoline), 8.35 (d, 1H, J=8.5 Hz, H4-quinoline), 8.37 (d, 1H,  $^3J=8.5$  Hz, H4-quinoline), 8.36 (s, 2H, H5, H7). ¹³C-NMR (50 MHz, DMSO):  $\delta$  70.66 (s), 74.38 (s), 114.50 (t), 116.44 (q), 117.74 (t), 119.45 (t), 120.08 (t), 121.1 (t), 123.89 (q), 125.91 (q), 126.39 (t), 126.42 (t), 126.95 (q), 127.61 (t), 127.73 (t), 128.32 (t), 128.41 (t), 129.40 (t), 129.58 (t), 129.69 (t), 130.99 (t), 131.16 (q), 136.53 (t), 136.82 (t), 137.00 (t), 139.76 (q), 146.55 (q), 146.68 (q), 151.53 (q), 154.59 (q), 156.54 (q), 156.82 (q), 157.85 (q) 163.73 (q), 172.28 (q). Anal.  $C_{36}H_{23}BrN_2O_6 \cdot 0.8HCl$ : C, H, N, Br.

5.2.2.5. 3-(2-Phthalimidoethoxy)-4'-[(2-quinolinyl)methoxy]-flavone-8-carboxylic acid **6e**: Mp 170.2–173.3 °C, yield 89%. 
¹H-NMR (400 MHz, DMSO):  $\delta$  3.54–3.55 (m, 2H, NCH<sub>2</sub>CH<sub>2</sub>O), 4.18–4.19 (m, 2H, NCH<sub>2</sub>CH<sub>2</sub>O), 5.44 (s, 2H, CH<sub>2</sub>O), 7.20 (d, 2H,  $^3J$  = 8.6 Hz, H3', H5'), 7.48–7.55 (m, 3H, H6, H5-phthalimide, H6-phthalimide), 7.61–7.65 (m, 1H, H6-quinoline), 7.68–7.70 (m, 2H, H4-phthalimide, H7-phthalimide), 7.80 (d, 1H,  $^3J$  = 8.5 Hz, H3-quinoline), 7.78–7.82 (m, 1H, H7-quinoline), 7.99–8.05 (m, 2H, H5-quinoline, H8-quinoline), 8.22–8.27 (m, 2H, H5, H7), 8.35 (d, 2H,  $^3J$  = 8.6 Hz, H2', H6'), 8.43 (d, 1H,  $^3J$  = 8.5 Hz, H4-quinoline).  $^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  69.92 (s), 70.67 (s), 70.68 (s), 114.75 (t), 119.34 (t), 122.13 (q), 122.89 (q), 124.05 (q), 124.16 (t), 126.43 (t), 126.97 (q), 127.29 (t), 127.74 (t), 128.31 (t), 128.97 (t), 129.70 (t), 130.45 (t), 130.80 (t), 135.49 (t), 136.93 (t) 137.85 (q), 138.75 (q), 146.69 (q), 152.36 (q), 154.39 (q), 155.74 (q), 160.01 (q), 165.12 (q), 167.71 (q), 168.43 (q), 173.22 (q). Anal.  $C_{36}$ H<sub>24</sub>N<sub>2</sub>O<sub>8</sub>·1.0HCl: C, H, N.

5.2.2.6. 3-[2-(N,N-Dimethylamino)ethoxy]-4'-(2-quinolinylmethoxy)flavone-8-carboxylic acid **6g**: A solution of 0.18 g (0.29 mmol) **6e** in 60 mL ethanol was heated under reflux for 1 h together with 10 mL hydrazine (1.1 equiv). The precipitate

(6f) was filtered. The crude amine (70 mg) was stirred overnight together with 0.5 mL 37% formaldehyde and 10 mg sodium cyanoborohydride at room temperature. The solvents were removed, the crude product was dissolved in ethanol. A few drops of 1 N HCl were added and the salt (35 mg) was obtained after filtration of the precipitate. Yield 21%, mp > 300 °C. <sup>1</sup>H-NMR (200 MHz, DMSO): δ 2.94 (d, 6H, NCH<sub>3</sub>), 3.48–3.53 (m, 2H, NCH<sub>2</sub>CH<sub>2</sub>O), 4.28–4.36 (m, 2H, NCH<sub>2</sub>CH<sub>2</sub>O), 5.36 (s, 2H, quinoline CH<sub>2</sub>O), 7.31 (d, 2H,  $^{3}J =$ 9.0 Hz, H3', H5'), 7.57 (t, 1H,  $^{3}J = 7.7$  Hz, H6), 7.68–7.75 (m, 1H, H6-quinoline), 7.83-7.93 (m, 2H, H3-quinoline, H7-quinoline), 8.16 (d, 1H,  $^{3}J = 9.3$  Hz, H8-quinoline), 8.28–8.35 (m, 5H, H2', H6', H5, H7, HCl), 8.66 (d, 1H,  $^{3}J$  = 8.5 Hz, H4quinoline), 10.99 (br s, 1H, ArCO<sub>2</sub>H). <sup>13</sup>C-NMR (50 MHz, DMSO): δ 42.18 (p), 55.54 (s), 65.51 (s), 69.81 (s), 115.05 (t), 119.63 (t), 121.56 (q), 122.60 (q), 124.01 (q), 124.52 (t), 126.87 (t), 127.06 (t), 127.14 (q), 128.02 (t), 129.07 (t), 130.51 (t), 130.67 (t), 135.87 (t), 138.18 (q), 138.73 (t), 145.03 (q), 152.48 (q), 155.04 (q), 156.35 (q), 160.11 (q), 173.21 (q).

5.2.3. 1-(5-Bromo-3-ethoxycarbonyl-2-hydroxylphenyl)-3-(3,5-dimethoxyphenyl)propane-1,3-dione **9** 

A solution of 10.0 g (35.0 mmol) ethyl 3-acetyl-5-bromo-2-hydroxybenzoate 8 and 11.7 g (55.2 mmol) methyl 3,5-dimethoxybenzoate 7 in 100 mL dry dioxane was added dropwise to a suspension of 6.0 g 60% NaH in 80 mL dioxane. The reaction mixture was heated under reflux overnight, and the bulk of dioxane was removed in vacuo. Petroleum ether 40–60 °C (400 mL) was added, and the precipitate was filtered. The crude product was suspended in water and acidified with 3 M HCl. The yellow precipitate was filtered and used in the next step without further purification.

5.2.4. 6-Bromo-3',5'-dimethoxyflavone-8-carboxylic acid 10

A solution of the diketone  $\ref{9}$  (16.62 g) in ethanol containing 2% of concentrated HCl was refluxed for 7 h. The bulk of ethanol was removed under reduced pressure, and the precipitate was collected by filtration. Recrystallisation from DMF yielded 6.06 g (15.0 mmol) of the pure product  $\ref{10}$ . Mp 207.3–207.8 °C, yield 43%.  $\ref{14}$ -NMR (200 MHz, DMSO):  $\ref{10}$  3.86 (s, 6H, 2 OCH<sub>3</sub>), 6.75 (bs, 1H, H4'), 7.33–7.36 (m, 3H, H3, H2', H6'), 8.33–8.37 (m, 2H, H5, H7). MS ( $\ref{10}$ /z): 405 [M + H]+ for  $\ref{10}$ C<sub>18</sub>H<sub>13</sub>BrO<sub>6</sub>.

5.2.5. 6-Bromo-3',5'-dihydroxyflavone-8-carboxylic acid 11

A solution (1 M) of BBr<sub>3</sub> in dichloromethane (88 mL, 88 mmol) was added dropwise to a suspension of **10** (5.77 g, 14.3 mmol) in 150 mL of dichloromethane. The reaction mixture was stirred at room temperature for 3 days, and poured out onto cracked ice and concentrated HCl. The yellow precipitate was collected by filtration, and washed with ethanol and petroleum ether 40–60 °C, yielding 4.98 g (13.2 mmol) of the dihydroxyflavone **11**. Mp > 300 °C, yield 93%. <sup>1</sup>H-NMR (200 MHz, DMSO):  $\delta$  6.51 (s, 1H, H3), 6.93 (s, 1H, H4), 6.99 (d, 2H,  $^4J$  = 2.1 Hz, H2', H5'), 8.29 (d, 2H,  $^4J$  = 2.2 Hz, H5, H7). <sup>13</sup>C-NMR (50 MHz, DMSO):  $\delta$  104.67 (t), 106.10 (t), 106.28 (t), 116.73 (q), 124.20 (q), 125.59 (q), 130.62 (t), 132.17 (q), 137.64 (t), 152.42 (q), 158.61 (2q), 163.17 (q), 163.67 (q), 175.06 (q). MS ( $^{\prime\prime\prime}$ ): 377 [M + H]+ for C<sub>16</sub>H<sub>9</sub>BrO<sub>6</sub>.

5.2.6. Ethyl 6-bromo-3',5'-dihydroxyflavone-8-carboxylate 12
A solution of 4.6 g (12.2 mmol) of 11 in 200 mL ethanol and 10 mL concentrated sulphuric acid was heated under reflux for 5 h. The reaction mixture was cooled to 4 °C, and the precipi-

tate was collected by filtration. Recrystallisation from DMF yielded 2.04 g (5.0 mmol) of pure compound 12. Mp > 300 °C, yield 41%. ¹H-NMR (200 MHz, DMSO):  $\delta$  1.40 (t, 3H,  $^3J$  = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 4.44 (q, 2H,  $^3J$  = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 6.49 (t, 1H,  $^4J$  = 2.0 Hz, H4'), 6.90 (s, 1H, H3), 6.95 (d, 2H,  $^4J$  = 2.0 Hz, H2', H6'), 8.28 (d, 1H,  $^4J$  = 2.6 Hz, H5/H7), 8.29 (d, 1H,  $^4J$  = 2.6 Hz, H5/H7), 9.73 (s, 2H, ArOH).  $^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  13.76 (p), 61.83 (s), 104.58 (t), 106.11 (t), 106.39 (t), 116.72 (q), 122.83 (q), 125.62 (q), 131.18 (t), 132.09 (t), 137.78 (t), 152.39 (q), 158.64 (q), 162.22 (q), 163.14 (q), 174.90 (q). MS (m/z): 405 [M + H]+ for C<sub>18</sub>H<sub>13</sub>BrO<sub>6</sub>.

# 5.2.7. Ethyl 6-bromo-3',5'-bis(2-quinolinemethoxy)flavone-8-carboxylate 13

A solution of 12 (1.93 g, 4.8 mmol), 2-chloromethyl quinoline hydrochloric acid (1.05 g, 4.8 mmol) and potassium carbonate (1.45 g, 10.5 mmol) in 30 mL DMF was stirred over night at 80 °C. The DMF was removed under reduced pressure, and water was added. The crude product was extracted with ethyl acetate, and the combined organic layers were dried over sodium sulphate. After the removal of the solvent, the residue was crystallised from DMF yielding a mixture of two products, which were separated with Amberlite resin IR-45(OH). The neutral product was characterised as the dialkylated flavone 13 (0.57 g, 0.83 mmol). Mp 189.1–189.2 °C, yield 17%. ¹H-NMR (200 MHz, DMSO):  $\delta$  1.39 (t, 3H,  $^3J$  = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 4.42 (q, 2H,  ${}^{3}J = 7.0 \text{ Hz}$ , OCH<sub>2</sub>CH<sub>3</sub>), 5.49 (s, 4H, 2 x OCH<sub>2</sub>), 7.10 (bs, 1H, H4'), 7.34 (s, 1H, H3), 7.57–7.72 (m, 6H, 2 x H6quinoline, 2 x H3-quinoline, H2', H6'), 7.77–7.83 (m, 2H,  ${}^{3}J$  = 7.6 Hz, 2 x H7-quinoline), 7.98–8.04 (m, 4H, 2 x H5-quinoline, 2 x H8-quinoline), 8.30 (d, 1H,  $^4J$  = 2.6 Hz, H5/7), 8.35 (d, 1H,  $^4J = 2.6$  Hz, H5/H7), 8.42 (d, 2H,  $^3J = 8.6$  Hz, H4-quinoline). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 14.16 (p), 61.95 (s), 71.33 (s), 105.31 (t), 106.26 (t), 107.27 (t), 117.67 (q), 118.97 (t), 122.47 (q), 125.94 (q), 126.55 (t), 127.40 (q), 127.53 (t), 128.72 (t), 129.81 (t), 132.59 (t), 132.96 (q), 137.07 (t), 138.87 (t), 147.21 (q), 153.11 (q), 156.71 (q), 159.96 (q), 162.37 (q), 163.03 (q), 176.02 (q). MS (m/z): 687 [M + H]+ for C38H27BrN2O6.

# 5.2.8. 6-Bromo-3',5'-bis(2-quinolinemethoxy)flavone-8-carbo-xylic acid 14

A solution of **13** (0.34 g, 0.49 mmol) in 30 mL DMF was heated to 60 °C, and 25 mL of 10% NaOH in water was then added. After stirring for 1 h, the reaction mixture was acidified with 3 M HCl and the precipitate was collected by filtration. Recrystallisation from DMF/ethanol yielded 0.24 g (0.36 mmol) of the pure product **14** as a white solid. Mp > 300 °C, yield 74%. ¹H-NMR (200 MHz, DMSO):  $\delta$  5.46 (s, 4H, OCH<sub>2</sub>), 7.01 (s, 1H, H4'), 7.20 (s, 1H, H3), 7.57–7.81 (m, 8H, 2 H3-quinoline, 2 x H6-quinoline, 2 x H7-quinoline, H2', H6'), 7.94–8.03 (m, 6H, 2 x H5-quinoline, 2 x H8-quinoline, H5, H7), 8.39 (d, 2H,  $^3J$  = 8.5 Hz, 2 x H4-quinoline).  $^{13}$ C-NMR (50 MHz, DMSO):  $\delta$  70.96 (s), 105.20 (t), 105.89 (t), 106.13 (t), 116.91 (q), 119.70 (t), 125.00 (q), 125.49 (t), 126.40 (t), 126.94 (q), 127.71 (t), 128.35 (t), 129.63 (t), 133.07 (q), 136.14 (t), 136.80 (t), 146.71 (q), 151.49 (q), 156.81 (q), 159.49 (q), 161.89 (q), 164.48 (q), 176.11 (q). Anal.  $C_{36}H_{23}BrN_2O_6$ -2HCl: C, H, N, Br.

### Acknowledgements

Kowa Company Ltd., Japan, is gratefully acknowledged for financial support of this research program. We thank Dr. F.J.J. de Kanter for the assistance with NMR-spectroscopy and Dr. B. van Baar for the collection of the mass spectra.

#### References

- [1] Samuelsson B., Science 220 (1983) 586-575.
- [2] Mayatepek E., Hoffmann G.F., Pediatr. Res. 37 (1995) 1–9.
- [3] Taylor I.K., Thorax 50 (1995) 1005-1010.
- [4] Pauwels R.A., Joos G.F., Kips J.C., Allergy 30 (1995) 615–622.
- [5] Ford-Hutchinson A.W., Adv. Prostaglandin Thromboxane Leukotriene Res. 23 (1995) 69–74.
- [6] Dahlen B., Dahlen S.E., Clin. Exp. Allergy 25 (1995) 50-54.
- [7] Griffin M., Weiss J.W., Leitch A.G., McFadden E.R., Corey E.J., Austen K.F., Drazen J.M., New Engl. J. Med. 308 (1983) 436.
- [8] Lam S., Chan H., LeRiche J.C., Chan-Yeung M., Salari H., J. Allergy Clin, Immunol. 81 (1988) 711–717.
- [9] Okubo T., Takahashi H., Sumitomo M., Shindoh K., Suzuki S., Int. Arch. Allergy Appl. Immun. 84 (1987) 149–155.
- [10] Brooks C.D.W., Summers J.B., J. Med. Chem. 39 (1996) 2629–2654.
- [11] Drazen J., Allergy 50 (1995) 22-26.
- [12] Fischer A.R., McFadden C.A., Frantz R., Awni W.M., Cohn J., Drazen J.M., Israel E., Am. J. Respir. Crit. Care Med. 152 (1995) 1203–1207.
- [13] Metters K.M., J. Lipid Mediators Cell Signalling 12 (1995) 413-427.
- [14] Gorenne I., Norel X., Brink C., Trends Pharmacol. Sci. 17 (1996) 342–345.
- [15] Zhang M.Q., Wada Y., Sato F., Timmerman H., J. Med. Chem. 38 (1995) 2472–2477.
- Zwaagstra M.E., Timmerman H., Abdoelgafoer R.S., Zhang M.Q., Eur. J. Med. Chem. 31 (1996) 861–874.
   Zwaagstra M.E., Timmerman H., Tamura M., Tohma T., Wada Y.,
- [17] Zwaagstra M.E., Timmerman H., Tamura M., Tohma T., Wada Y. Onogi K., Zhang M.Q., J. Med. Chem. 40 (1997) 1075–1089.
- [18] Zhang M.Q., Zwaagstra M.E., Current Med. Chem. 4 (1997) 229-246.
- [19] Zhang M.Q., van de Stulpe A., Zuiderveld O.P., Timmerman H., Eur. J. Med. Chem. 32 (1997) 95–102.
- [20] Zhang M.Q., Zwaagstra M.E., Nederkoorn P.H.J., Timmerman H., Bioorg. Med. Chem. Lett. 7 (1997) 1331–1336.
- [21] Algar J., Flynn J.J., Proc. Roy. Irish Acad. 42B (1934) 1-2.
- [22] Oyamada T.J., Chem. Soc. Jpn. 55 (1934) 1256-1259
- [23] Borch R.F., Hassid A.I., J. Org. Chem. 37 (1972) 1673–1674.
- [24] Wu E.S.C., Cole T.E., Davidson T.A., Dailey M.A., Doring K.G., Fedorchuk M., Loch J.T., Thomas T.L., Blosser F.C., Borrelli A.R., Kinsolving C.R., Parker R.B., Strand J.C., Watkins B.E., J. Med. Chem. 32 (1989) 183–192.
- [25] Erickson R.H., Natalie K.J., Bock W., Lu Z., Farzin F., Sherill R.G., Meloni D.J., Patch R., Rzesotarski W.J., Clifton J., Pontecorvo M.J., Bailey M.A., Naper K., Karbon W., J. Med. Chem. 35 (1992) 1526–1535.
- [26] Ares J.J., Outt P.E., Kakodkar S.V., Buss R.C., Geiger J.C., J. Org. Chem. 58 (1993) 7903–7905.
- [27] Zhang M.Q., Leurs R., Timmerman H., in: Wolff M.E. (Ed.), Burger's Medicinal Chemistry and Drug Discovery, Vol. 5: Therapeutic Agents (5th ed.), Wiley, New York, 1997, pp. 495–559.
- [28] Adams G.K., Lichtenstein L., J. Immunol. 122 (1979) 555-562.
- [29] Eiser N., Denman W., Hayhurst M., Ancic P., Rodriguez J., Thorax 44 (1989) 334–335.
- [30] Hedqvist P., Lindbom L., Palmertz U., Raud J., Adv. Prostaglandin Thromboxane Leukotriene Res. 22 (1994) 91–99.
- [31] Zhang M.Q., Terlaak A.M., Timmerman H., Eur. J. Med. Chem. 28 (1993) 165-173.